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ТҮВДЭНЖАМЦЫН БАЛЖИННЯМ

**TLR9/ IFN-γ-ЫН ХАРИЛЦАН ДЭМЖИХ ҮЙЛЧЛЭЛД ЭЕРЭГ БОЛОН
СӨРӨГ ЗОХИЦУУЛАГЧ УУРГИЙН ИДЭВХЖЛИЙГ ҮНЭЛСЭН ДҮН**

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ТОВЧ ХУРААНГУЙ

Сэдэв: TLR9/IFN- γ -ийн харилцан дэмжих үйлчлэлд эерэг болон сөрөг зохицуулагч уургийн идэвхжлийг үнэлсэн дүн

Үндэслэл: Анхдагч дархлааны TLR9 нь бактери, вирус, эукариот биетийн доторх метилжээгүй CpG ДНХ-ийн дарааллыг таньснаар дархлааны урьтал цитокин болон IFN- α , IFN- β -ын нийлэгжлийг өдөөж дархлааны урвалыг эхлүүлдэг. Өвөрмөц дархлаа тогтолцооны IFN- γ нь дархлааны үндсэн эсүүдээс ялгарч халдварын эсрэг өвөрмөц, хүчтэй урвалыг өрнүүлдэг. Бидний өмнөх судалгаагаар TLR9-ийн лиганд CpG ДНХ IFN- γ -ийн дохио дамжилтыг нэмэгдүүлж байсан. TLR9/IFN- γ -ийн харилцан дэмжих үйлчлэлийг нөхцөлдүүлэгч гол уургийг илрүүлэх нь энэ судалгааны үндэслэл боллоо.

Зорилго: IFN- γ /TLR9-ийн харилцан дэмжих үйлчлэлд эерэг болон сөрөг зохицуулагч уургуудын идэвхжлийг тодорхойлох

Материал, арга аргачлал: Энэхүү судалгааг АШУҮИС-ийн ШУТГ-ийн харьяа Цөм лабораторид хийж гүйцэтгэсэн. Судалгаанд хулганын аортын шугаман эндотель эсийг (END-D) өсгөвөрийг ашигласан. Эндотель эсийг TLR9-ийн лиганд CpG ДНХ өдөөсөөс 1 цагын дараа IFN- γ -аар үйлчилсэн. IFN- γ -аар үйлчилсэнээс 0.5, 1, 2, 4, 8 цагийн дараа эсийг задалж уургийг ялгасан. Иммуноблоттингийн шинжилгээгээр сөрөг (SHP-2, SOCS1, PIAS1) болон эерэг (p38) зохицуулагч уургийн нийлэгжлийг тодорхойлсон.

Үр дүн: TLR9-ийн лиганд CpG ДНХ-ээр өдөөхөд IFN- γ -ийн эерэг зохицуулагч p38 уургийн фосфоржилтыг 1-2 цагт эрс нэмэгдүүлсэн. CpG ДНХ нь IFN- γ -ийн сөрөг зохицуулагч SOCS1, PIAS1 уургийн нийлэгжлийг 2-8 цагуудад бууруулж байсан. Харин SHP-2 уургийн нийлэгжилд нөлөө үзүүлээгүй.

Дүгнэлт: TLR9-ийн лиганд CpG ДНХ нь IFN- γ -ийн сөрөг зохицуулагч SOCS1, PIAS1 уургийн нийлэгжлийг бууруулж байна. CpG ДНХ нь IFN- γ -ийн эерэг зохицуулагч p-38 уургийн фосфоржилтыг нэмэгдүүлж байна. Therefore, TLR9 ligand CpG DNA increased IFN- γ signal transduction in endothelial cell line.

Түлхүүр үг: CpG DNA, p38, PIAS1, SOCS1, SHP2, END-D

ABSTRACT

Title: Result of negative and positive regulatory protein activation in the TLR9/IFN- γ synergistic signal transduction

Introduction: TLR9 recognizes unmethylated CpG DNA of bacteria, virus, and intracellular parasite and induce IFN- α , IFN- β and pro-inflammatory cytokine expression which results in the activation of immune response. IFN- γ produced from immune cell has a strong effect against an infection. In our previous study, we revealed that there is a synergistic action between TLR9 and IFN- γ signaling in the endothelial cells. We performed the experiment in order to determine a protein that induces synergistic action of TLR9 ligand CpG DNA/IFN- γ signal transduction.

Purpose: To determine the role of negative and positive regulator proteins on the IFN- γ /TLR9 synergistic signaling pathway.

Materials and Methods: This study was held in the Core Laboratory, Science Technology Center, Mongolian National University of Medical Sciences (MNUMS). In this study, murine endothelial cell (END-D) culture was used. END-D cells were seeded in a 6-well plastic plate, pre-treated with TLR9 ligand CpG DNA and then stimulated with IFN- γ . The negative (SHP-2, SOCS1, PIAS1) and positive (p38) regulator protein expression was detected by Western blotting.

Results: We assessed viability of the cells that treated with up to 20 μ M of CpG DNA, using CCK 8 kit. Treatment by TLR9 ligand CpG DNA and IFN- γ increased positive regulator p38 phosphorylation at 1-2 hours. CpG DNA inhibited IFN- γ negative regulator SOCS1 and PIAS1 protein expression at 2-8 hours and SHP-2 expression could not affect. Therefore, TLR9 ligand CpG DNA increased IFN- γ signal transduction in endothelial cell line.

Conclusion: TLR9 ligand CpG DNA has decreased IFN- γ negative regulator protein PIAS1 and SOCS1 expression. CpG DNA has increased IFN- γ positive regulator protein p38 phosphorylation.

Keyword: CpG DNA, p38, PIAS1, SOCS1, SHP2, END-D

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1. Dalpke AH, Eckerle S, Frey M, Heeg K. Triggering of Toll-like receptors modulates IFN-gamma signaling: involvement of serine 727 STAT1 phosphorylation and suppressors of cytokine signaling. *Eur J Immunol*. Jul 2003;33(7):1776-1787.
2. Ohto U, Shibata T, Tanji H, et al. Structural basis of CpG and inhibitory DNA recognition by Toll-like receptor 9. *Nature*. Apr 30 2015;520(7549):702-705.
3. Blouin CM, Hamon Y, Gonnord P, et al. Glycosylation-Dependent IFN-gammaR Partitioning in Lipid and Actin Nanodomains Is Critical for JAK Activation. *Cell*. Aug 11 2016;166(4):920-934.
4. Isaacs A, Lindenmann J. Virus interference. I. The interferon. *Proceedings of the Royal Society of London. Series B, Biological sciences*. Sep 12 1957;147(927):258-267.
5. Fitzner N, Zahner L, Habich C, Kolb-Bachofen V. Stimulatory type A CpG-DNA induces a Th2-like response in human endothelial cells. *International immunopharmacology*. Nov 2011;11(11):1789-1795.
6. Krieg AM, Vollmer J. Toll-like receptors 7, 8, and 9: linking innate immunity to autoimmunity. *Immunological reviews*. Dec 2007;220:251-269.
7. Fensterl V, Sen GC. Interferons and viral infections. *BioFactors (Oxford, England)*. Jan-Feb 2009;35(1):14-20.
8. Vilcek J. Novel interferons. *Nature immunology*. Jan 2003;4(1):8-9.
9. Sadir R, Forest E, Lortat-Jacob H. The heparan sulfate binding sequence of interferon-gamma increased the on rate of the interferon-gamma-interferon-gamma receptor complex formation. *The Journal of biological chemistry*. May 1 1998;273(18):10919-10925.
10. Lortat-Jacob H, Grimaud JA. Interferon-gamma binds to heparan sulfate by a cluster of amino acids located in the C-terminal part of the molecule. *FEBS letters*. Mar 11 1991;280(1):152-154.
11. Frucht DM, Fukao T, Bogdan C, Schindler H, O'Shea JJ, Koyasu S. IFN-gamma production by antigen-presenting cells: mechanisms emerge. *Trends in immunology*. Oct 2001;22(10):556-560.
12. Sen GC. Viruses and interferons. *Annual review of microbiology*. 2001;55:255-281.
13. Баярт Б, Түвшинжаргал Ц. 2015;6(2):ху:49.
14. Bazan JF. Structural design and molecular evolution of a cytokine receptor superfamily. *Proceedings of the National Academy of Sciences of the United States of America*. Sep 1990;87(18):6934-6938.
15. Thoreau E, Petridou B, Kelly PA, Djiane J, Mornon JP. Structural symmetry of the extracellular domain of the cytokine/growth hormone/prolactin receptor family and interferon receptors revealed by hydrophobic cluster analysis. *FEBS letters*. Apr 22 1991;282(1):26-31.
16. Kaplan DH, Greenlund AC, Tanner JW, Shaw AS, Schreiber RD. Identification of an interferon-gamma receptor alpha chain sequence required for JAK-1 binding. *The Journal of biological chemistry*. Jan 05 1996;271(1):9-12.
17. Greenlund AC, Farrar MA, Viviano BL, Schreiber RD. Ligand-induced IFN gamma receptor tyrosine phosphorylation couples the receptor to its signal transduction system (p91). *The EMBO journal*. Apr 01 1994;13(7):1591-1600.
18. Farrar MA, Fernandez-Luna J, Schreiber RD. Identification of two regions within the cytoplasmic domain of the human interferon-gamma receptor required for function. *The Journal of biological chemistry*. Oct 15 1991;266(29):19626-19635.
19. Farrar MA, Campbell JD, Schreiber RD. Identification of a functionally important sequence in the C terminus of the interferon-gamma receptor. *Proceedings of the National Academy of Sciences of the United States of America*. Dec 15 1992;89(24):11706-11710.
20. Cook JR, Jung V, Schwartz B, Wang P, Pestka S. Structural analysis of the human interferon gamma receptor: a small segment of the intracellular domain is specifically required for class I major histocompatibility complex antigen induction and antiviral activity. *Proceedings of the National Academy of Sciences of the United States of America*. Dec 01 1992;89(23):11317-11321.

108. You M, Yu DH, Feng GS. Shp-2 tyrosine phosphatase functions as a negative regulator of the interferon-stimulated Jak/STAT pathway. *Molecular and cellular biology*. Mar 1999;19(3):2416-2424.
109. An H, Zhao W, Hou J, et al. SHP-2 phosphatase negatively regulates the TRIF adaptor protein-dependent type I interferon and proinflammatory cytokine production. *Immunity*. Dec 2006;25(6):919-928.
110. Park JH, Ko R, Lee SY. Reciprocal regulation of TLR2-mediated IFN-beta production by SHP2 and Gsk3beta. *Scientific reports*. Jul 28 2017;7(1):6807.
111. Kim EJ, Lee SM, Suk K, Lee WH. CD300a and CD300f differentially regulate the MyD88 and TRIF-mediated TLR signalling pathways through activation of SHP-1 and/or SHP-2 in human monocytic cell lines. *Immunology*. Mar 2012;135(3):226-235.
112. Teng JF, Wang K, Jia ZM, et al. Lentivirus-Mediated Silencing of Src Homology 2 Domain-Containing Protein Tyrosine Phosphatase 2 Inhibits Release of Inflammatory Cytokines and Apoptosis in Renal Tubular Epithelial Cells Via Inhibition of the TLR4/NF-kB Pathway in Renal Ischemia-Reperfusion Injury. *Kidney & blood pressure research*. 2018;43(4):1084-1103.
113. Liu B, Shuai K. Induction of apoptosis by protein inhibitor of activated Stat1 through c-Jun NH2-terminal kinase activation. *The Journal of biological chemistry*. Sep 28 2001;276(39):36624-36631.
114. Coccia EM, Stellacci E, Orsatti R, et al. Protein inhibitor of activated signal transducer and activator of transcription (STAT)-1 (PIAS-1) regulates the IFN-gamma response in macrophage cell lines. *Cellular signalling*. Jun 2002;14(6):537-545.
115. Brown JR, Conn KL, Wasson P, et al. SUMO Ligase Protein Inhibitor of Activated STAT1 (PIAS1) Is a Constituent Promyelocytic Leukemia Nuclear Body Protein That Contributes to the Intrinsic Antiviral Immune Response to Herpes Simplex Virus 1. *Journal of virology*. Jul 1 2016;90(13):5939-5952.

21. Kotenko SV, Izotova LS, Pollack BP, et al. Interaction between the components of the interferon gamma receptor complex. *The Journal of biological chemistry*. Sep 08 1995;270(36):20915-20921.
22. Bach EA, Tanner JW, Marsters S, et al. Ligand-induced assembly and activation of the gamma interferon receptor in intact cells. *Molecular and cellular biology*. Jun 1996;16(6):3214-3221.
23. Farrar MA, Schreiber RD. The molecular cell biology of interferon-gamma and its receptor. *Annual review of immunology*. 1993;11:571-611.
24. Pestka S, Kotenko SV, Muthukumaran G, Izotova LS, Cook JR, Garotta G. The interferon gamma (IFN-gamma) receptor: a paradigm for the multichain cytokine receptor. *Cytokine & growth factor reviews*. Sep 1997;8(3):189-206.
25. Villarino AV, Kanno Y, O'Shea JJ. Mechanisms and consequences of Jak-STAT signaling in the immune system. *Nature immunology*. Mar 22 2017;18(4):374-384.
26. Braunstein J, Brutsaert S, Olson R, Schindler C. STATs dimerize in the absence of phosphorylation. *The Journal of biological chemistry*. Sep 05 2003;278(36):34133-34140.
27. Wenta N, Strauss H, Meyer S, Vinkemeier U. Tyrosine phosphorylation regulates the partitioning of STAT1 between different dimer conformations. *Proceedings of the National Academy of Sciences of the United States of America*. Jul 8 2008;105(27):9238-9243.
28. Chen X, Xu H, Yuan P, et al. Integration of external signaling pathways with the core transcriptional network in embryonic stem cells. *Cell*. Jun 13 2008;133(6):1106-1117.
29. Willi M, Yoo KH, Wang C, Trajanoski Z, Hennighausen L. Differential cytokine sensitivities of STAT5-dependent enhancers rely on Stat5 autoregulation. *Nucleic acids research*. Dec 01 2016;44(21):10277-10291.
30. Ciofani M, Madar A, Galan C, et al. A validated regulatory network for Th17 cell specification. *Cell*. Oct 12 2012;151(2):289-303.
31. Qing Y, Stark GR. Alternative activation of STAT1 and STAT3 in response to interferon-gamma. *The Journal of biological chemistry*. Oct 01 2004;279(40):41679-41685.
32. Yang XP, Ghoreschi K, Steward-Tharp SM, et al. Opposing regulation of the locus encoding IL-17 through direct, reciprocal actions of STAT3 and STAT5. *Nature immunology*. Mar 2011;12(3):247-254.
33. Hirahara K, Onodera A, Villarino AV, et al. Asymmetric Action of STAT Transcription Factors Drives Transcriptional Outputs and Cytokine Specificity. *Immunity*. May 19 2015;42(5):877-889.
34. Veals SA, Schindler C, Leonard D, et al. Subunit of an alpha-interferon-responsive transcription factor is related to interferon regulatory factor and Myb families of DNA-binding proteins. *Molecular and cellular biology*. Aug 1992;12(8):3315-3324.
35. Watford WT, Hissong BD, Durant LR, et al. Tpl2 kinase regulates T cell interferon-gamma production and host resistance to *Toxoplasma gondii*. *The Journal of experimental medicine*. Nov 24 2008;205(12):2803-2812.
36. Shultz DB, Fuller JD, Yang Y, Sizemore N, Rani MR, Stark GR. Activation of a subset of genes by IFN-gamma requires IKKbeta but not interferon-dependent activation of NF-kappaB. *Journal of interferon & cytokine research : the official journal of the International Society for Interferon and Cytokine Research*. Oct 2007;27(10):875-884.
37. Stark GR, Darnell JE, Jr. The JAK-STAT pathway at twenty. *Immunity*. Apr 20 2012;36(4):503-514.
38. Begitt A, Droscher M, Meyer T, et al. STAT1-cooperative DNA binding distinguishes type 1 from type 2 interferon signaling. *Nature immunology*. Feb 2014;15(2):168-176.
39. Akira S, Takeda K. Toll-like receptor signalling. *Nat Rev Immunol*. Jul 2004;4(7):499-511.
40. Schindler H, Lutz MB, Rollinghoff M, Bogdan C. The production of IFN-gamma by IL-12/IL-18-activated macrophages requires STAT4 signaling and is inhibited by IL-4. *Journal of immunology*. Mar 01 2001;166(5):3075-3082.
41. Fukao T, Frucht DM, Yap G, Gadina M, O'Shea JJ, Koyasu S. Inducible expression of Stat4 in dendritic cells and macrophages and its critical role in innate and adaptive immune responses. *Journal of immunology*. Apr 01 2001;166(7):4446-4455.

42. Hochrein H, Shortman K, Vremec D, Scott B, Hertzog P, O'Keeffe M. Differential production of IL-12, IFN-alpha, and IFN-gamma by mouse dendritic cell subsets. *Journal of immunology*. May 01 2001;166(9):5448-5455.
43. Liu B, Mink S, Wong KA, et al. PIAS1 selectively inhibits interferon-inducible genes and is important in innate immunity. *Nature immunology*. Sep 2004;5(9):891-898.
44. Kubo M, Hanada T, Yoshimura A. Suppressors of cytokine signaling and immunity. *Nature immunology*. Dec 2003;4(12):1169-1176.
45. Alexander WS, Hilton DJ. The role of suppressors of cytokine signaling (SOCS) proteins in regulation of the immune response. *Annual review of immunology*. 2004;22:503-529.
46. Yoshimura A, Naka T, Kubo M. SOCS proteins, cytokine signalling and immune regulation. *Nature reviews. Immunology*. Jun 2007;7(6):454-465.
47. Cassel SL, Rothman PB. Chapter 3: Role of SOCS in allergic and innate immune responses. *Advances in immunology*. 2009;103:49-76.
48. Shao RX, Zhang L, Hong Z, et al. SOCS1 abrogates IFN's antiviral effect on hepatitis C virus replication. *Antiviral research*. Feb 2013;97(2):101-107.
49. Kamizono S, Hanada T, Yasukawa H, et al. The SOCS box of SOCS-1 accelerates ubiquitin-dependent proteolysis of TEL-JAK2. *The Journal of biological chemistry*. Apr 20 2001;276(16):12530-12538.
50. Ryo A, Suizu F, Yoshida Y, et al. Regulation of NF-kappaB signaling by Pin1-dependent prolyl isomerization and ubiquitin-mediated proteolysis of p65/RelA. *Molecular cell*. Dec 2003;12(6):1413-1426.
51. Fujimoto M, Naka T. SOCS1, a Negative Regulator of Cytokine Signals and TLR Responses, in Human Liver Diseases. *Gastroenterology research and practice*. 2010;2010.
52. Kinjyo I, Hanada T, Inagaki-Ohara K, et al. SOCS1/JAB is a negative regulator of LPS-induced macrophage activation. *Immunity*. Nov 2002;17(5):583-591.
53. Nakagawa R, Naka T, Tsutsui H, et al. SOCS-1 participates in negative regulation of LPS responses. *Immunity*. Nov 2002;17(5):677-687.
54. Naka T, Fujimoto M, Tsutsui H, Yoshimura A. Negative regulation of cytokine and TLR signalings by SOCS and others. *Advances in immunology*. 2005;87:61-122.
55. Shuai K, Liu B. Regulation of JAK-STAT signalling in the immune system. *Nature reviews. Immunology*. Nov 2003;3(11):900-911.
56. Rytinki MM, Kaikkonen S, Pehkonen P, Jaaskelainen T, Palvimo JJ. PIAS proteins: pleiotropic interactors associated with SUMO. *Cellular and molecular life sciences : CMLS*. Sep 2009;66(18):3029-3041.
57. Aravind L, Koonin EV. SAP - a putative DNA-binding motif involved in chromosomal organization. *Trends in biochemical sciences*. Mar 2000;25(3):112-114.
58. Shuai K, Liu B. Regulation of gene-activation pathways by PIAS proteins in the immune system. *Nature reviews. Immunology*. Aug 2005;5(8):593-605.
59. Duval D, Duval G, Keding C, Poch O, Boeuf H. The 'PINIT' motif, of a newly identified conserved domain of the PIAS protein family, is essential for nuclear retention of PIAS3L. *FEBS letters*. Nov 6 2003;554(1-2):111-118.
60. Hochstrasser M. SP-RING for SUMO: new functions bloom for a ubiquitin-like protein. *Cell*. Oct 5 2001;107(1):5-8.
61. Stehmeier P, Muller S. Phospho-regulated SUMO interaction modules connect the SUMO system to CK2 signaling. *Molecular cell*. Feb 13 2009;33(3):400-409.
62. Lee KM, O'Connell MJ. A new SUMO ligase in the DNA damage response. *DNA repair*. Jan 5 2006;5(1):138-141.
63. Jiang Y, Li Z, Schwarz EM, et al. Structure-function studies of p38 mitogen-activated protein kinase. Loop 12 influences substrate specificity and autophosphorylation, but not upstream kinase selection. *The Journal of biological chemistry*. Apr 25 1997;272(17):11096-11102.
64. Zarubin T, Han J. Activation and signaling of the p38 MAP kinase pathway. *Cell research*. Jan 2005;15(1):11-18.
65. Chen FF, Jiang G, Xu K, Zheng JN. Function and mechanism by which interferon regulatory factor-1 inhibits oncogenesis. *Oncology letters*. Feb 2013;5(2):417-423.

66. Lohoff M, Mak TW. Roles of interferon-regulatory factors in T-helper-cell differentiation. *Nature reviews. Immunology*. Feb 2005;5(2):125-135.
67. Dornan D, Eckert M, Wallace M, et al. Interferon regulatory factor 1 binding to p300 stimulates DNA-dependent acetylation of p53. *Molecular and cellular biology*. Nov 2004;24(22):10083-10098.
68. Negishi H, Fujita Y, Yanai H, et al. Evidence for licensing of IFN-gamma-induced IFN regulatory factor 1 transcription factor by MyD88 in Toll-like receptor-dependent gene induction program. *Proceedings of the National Academy of Sciences of the United States of America*. Oct 10 2006;103(41):15136-15141.
69. Fulda S, Debatin KM. IFN-gamma sensitizes for apoptosis by upregulating caspase-8 expression through the Stat1 pathway. *Oncogene*. Apr 4 2002;21(15):2295-2308.
70. Wang J, Zhang W, Zhang Y, et al. c-Jun N-terminal kinase (JNK1) upregulates XIAP-associated factor 1 (XAF1) through interferon regulatory factor 1 (IRF-1) in gastrointestinal cancer. *Carcinogenesis*. Feb 2009;30(2):222-229.
71. Nguyen H, Lin R, Hiscott J. Activation of multiple growth regulatory genes following inducible expression of IRF-1 or IRF/RelA fusion proteins. *Oncogene*. Sep 18 1997;15(12):1425-1435.
72. Park SY, Seol JW, Lee YJ, et al. IFN-gamma enhances TRAIL-induced apoptosis through IRF-1. *European journal of biochemistry*. Nov 2004;271(21):4222-4228.
73. Hansson GK, Edfeldt K. Toll to be paid at the gateway to the vessel wall. *Arterioscler Thromb Vasc Biol*. Jun 2005;25(6):1085-1087.
74. Ishii KJ, Akira S. Innate immune recognition of, and regulation by, DNA. *Trends in immunology*. Nov 2006;27(11):525-532.
75. Frazao JB, Errante PR, Condino-Neto A. Toll-like receptors' pathway disturbances are associated with increased susceptibility to infections in humans. *Archivum immunologiae et therapiae experimentalis*. Dec 2013;61(6):427-443.
76. Latz E, Visintin A, Espevik T, Golenbock DT. Mechanisms of TLR9 activation. *Journal of endotoxin research*. 2004;10(6):406-412.
77. Hemmi H, Takeuchi O, Kawai T, et al. A Toll-like receptor recognizes bacterial DNA. *Nature*. Dec 07 2000;408(6813):740-745.
78. Singer M, de Waaij DJ, Morré SA, Ouburg S. CpG DNA analysis of bacterial STDs. *BMC Infectious Diseases*. 07/1604/17/received07/08/accepted 2015;15:273.
79. Du X, Poltorak A, Wei Y, Beutler B. Three novel mammalian toll-like receptors: gene structure, expression, and evolution. *European cytokine network*. Sep 2000;11(3):362-371.
80. McKelvey KJ, Highton J, Hessian PA. Cell-specific expression of TLR9 isoforms in inflammation. *Journal of autoimmunity*. Feb 2011;36(1):76-86.
81. Gursel M, Verthelyi D, Gursel I, Ishii KJ, Klinman DM. Differential and competitive activation of human immune cells by distinct classes of CpG oligodeoxynucleotide. *Journal of leukocyte biology*. May 2002;71(5):813-820.
82. Mao TK, Lian ZX, Selmi C, et al. Altered monocyte responses to defined TLR ligands in patients with primary biliary cirrhosis. *Hepatology*. Oct 2005;42(4):802-808.
83. Azulay-Debby H, Edry E, Melamed D. CpG DNA stimulates autoreactive immature B cells in the bone marrow. *European journal of immunology*. Jun 2007;37(6):1463-1475.
84. Gurcan HM, Keskin DB, Stern JN, Nitzberg MA, Shekhani H, Ahmed AR. A review of the current use of rituximab in autoimmune diseases. *International immunopharmacology*. Jan 2009;9(1):10-25.
85. Buhe V, Pers JO, Marianowski R, Berthou C, Youinou P, Loisel S. Development of a Murine model to dissect the CpG-oligonucleotide-enhancement of the killing of human B Cells by rituximab. *Journal of autoimmunity*. Mar 2010;34(2):136-144.
86. Gay NJ, Gangloff M. Structure and function of Toll receptors and their ligands. *Annual review of biochemistry*. 2007;76:141-165.
87. Krieg AM, Yi AK, Matson S, et al. CpG motifs in bacterial DNA trigger direct B-cell activation. *Nature*. Apr 6 1995;374(6522):546-549.
88. Hu X, Ivashkiv LB. Cross-regulation of signaling pathways by interferon-gamma: implications for immune responses and autoimmune diseases. *Immunity*. Oct 16 2009;31(4):539-550.

89. Schroder K, Hertzog PJ, Ravasi T, Hume DA. Interferon-gamma: an overview of signals, mechanisms and functions. *Journal of leukocyte biology*. Feb 2004;75(2):163-189.
90. Chaudhry UI, Kingham TP, Plitas G, Katz SC, Raab JR, DeMatteo RP. Combined stimulation with interleukin-18 and CpG induces murine natural killer dendritic cells to produce IFN-gamma and inhibit tumor growth. *Cancer research*. Nov 01 2006;66(21):10497-10504.
91. Tsolmongyn B, Koide N, Jambalmaniin U, et al. A Toll-like receptor 2 ligand, Pam3CSK4, augments interferon-gamma-induced nitric oxide production via a physical association between MyD88 and interferon-gamma receptor in vascular endothelial cells. *Immunology*. Nov 2013;140(3):352-361.
92. Sumpio BE, Riley JT, Dardik A. Cells in focus: endothelial cell. *The international journal of biochemistry & cell biology*. Dec 2002;34(12):1508-1512.
93. Hardin C, Rajendran K, Manomohan G, et al. Glassy dynamics, cell mechanics, and endothelial permeability. *The journal of physical chemistry. B*. Oct 24 2013;117(42):12850-12856.
94. McCarthy CG, Wenceslau CF, Gouloupoulou S, Ogbi S, Matsumoto T, Webb RC. Autoimmune therapeutic chloroquine lowers blood pressure and improves endothelial function in spontaneously hypertensive rats. *Pharmacological research*. Nov 2016;113(Pt A):384-394.
95. Salvador B, Arranz A, Francisco S, et al. Modulation of endothelial function by Toll like receptors. *Pharmacological research*. Jun 2016;108:46-56.
96. Shahrakyvahed A, Sanchooli J, Sanadgol N, Arababadi MK, Kennedy D. TLR9: an important molecule in the fight against hepatitis B virus. *Postgraduate medical journal*. Jul 2014;90(1065):396-401.
97. Tatematsu M, Seya T, Matsumoto M. [RNA structure recognized by Toll-like receptor 3 in innate immunity]. *Seikagaku. The Journal of Japanese Biochemical Society*. Aug 2014;86(4):523-527.
98. Lees JR. Interferon gamma in autoimmunity: A complicated player on a complex stage. *Cytokine*. Jul 2015;74(1):18-26.
99. Wormald S, Zhang JG, Krebs DL, et al. The comparative roles of suppressor of cytokine signaling-1 and -3 in the inhibition and desensitization of cytokine signaling. *The Journal of biological chemistry*. Apr 21 2006;281(16):11135-11143.
100. Mori D, Koide N, Tsolmongyn B, et al. Poly I:C enhances production of nitric oxide in response to interferon-gamma via upregulation of interferon regulatory factor 7 in vascular endothelial cells. *Microvascular research*. Mar 2015;98:68-73.
101. Qin L, Huang Q, Zhang H, et al. SOCS1 prevents graft arteriosclerosis by preserving endothelial cell function. *Journal of the American College of Cardiology*. Jan 7-14 2014;63(1):21-29.
102. Tamiya T, Kashiwagi I, Takahashi R, Yasukawa H, Yoshimura A. Suppressors of cytokine signaling (SOCS) proteins and JAK/STAT pathways: regulation of T-cell inflammation by SOCS1 and SOCS3. *Arteriosclerosis, thrombosis, and vascular biology*. May 2011;31(5):980-985.
103. DiGiandomenico A, Wylezinski LS, Hawiger J. Intracellular delivery of a cell-penetrating SOCS1 that targets IFN-gamma signaling. *Science signaling*. Jul 21 2009;2(80):ra37.
104. Skjesol A, Liebe T, Iliev DB, et al. Functional conservation of suppressors of cytokine signaling proteins between teleosts and mammals: Atlantic salmon SOCS1 binds to JAK/STAT family members and suppresses type I and II IFN signaling. *Developmental and comparative immunology*. Jul 2014;45(1):177-189.
105. Recio C, Oguiza A, Lazaro I, Mallavia B, Egidio J, Gomez-Guerrero C. Suppressor of cytokine signaling 1-derived peptide inhibits Janus kinase/signal transducers and activators of transcription pathway and improves inflammation and atherosclerosis in diabetic mice. *Arteriosclerosis, thrombosis, and vascular biology*. Sep 2014;34(9):1953-1960.
106. Wang S, Zheng G, Zhao L, Xu F, Qian J. Shp-2 contributes to anti-RSV activity in human pulmonary alveolar epithelial cells by interfering with the IFN-alpha-induced Jak/Stat1 pathway. *Journal of cellular and molecular medicine*. Oct 2015;19(10):2432-2440.
107. Baron M, Davignon JL. Inhibition of IFN-gamma-induced STAT1 tyrosine phosphorylation by human CMV is mediated by SHP2. *Journal of immunology*. Oct 15 2008;181(8):5530-5536.